Selective Immunoglobulin M Deficiency in an Adult With *Streptococcus pneumoniae* Sepsis and Invasive Aspergillosis

R Hong, S Gupta

Division of Basic and Clinical Immunology, University of California, Irvine, California, USA

Abstract

Primary selective immunoglobulin (Ig) M deficiency usually presents early in life with recurrent or severe infections caused by encapsulated and gram-negative organisms. Primary selective IgM deficiency in adults is rare and is usually associated with autoimmune diseases or malignant neoplasm. We performed an extensive immunological analysis of innate and adaptive immunity in an adult patient with possible primary selective IgM deficiency who presented with life-threatening *Streptococcus pneumoniae* septic shock and invasive *Aspergillus fumigatus* infection. The patient had no evidence of autoimmune disease or malignant neoplasm. Serum IgG, IgA, and IgE were normal; however, serum IgM levels and specific antibody titers against all 14 pneumococcal polysaccharide serotypes were consistently low. Complement CH50, C3, C4, and neutrophil phagocytosis and oxidative burst were normal. Toll-like receptor expression on monocytes was also normal. Therefore, adult patients with serious life-threatening and unusual infections should be investigated for possible selective primary IgM deficiency.

Key words: Selective IgM deficiency. Primary immunodeficiency. Unusual infections. Toll-like receptors. Autoantibodies. Aspergillosis. Complement. Immune complexes.

Resumen

La insuficiencia de IgM selectiva primaria se manifiesta habitualmente en una edad temprana con infecciones recurrentes o graves causadas por bacterias gram-negativas encapsuladas. La insuficiencia de IgM selectiva primaria es muy poco frecuente en adultos y normalmente se asocia con enfermedades autoinmunes o neoplasias malignas. Se llevó a cabo un análisis inmunológico extensivo de la inmunidad innata y adaptativa en un paciente adulto con una posible insuficiencia de IgM selectiva primaria que presentó un choque septicémico de *Streptococcus pneumoniae* que podía poner en peligro su vida y una infección por *Aspergillus fumigatus* invasiva. El paciente no mostraba signos de ninguna enfermedad autoinmune ni de neoplasia maligna. La IgG, IgA e IgE séricas eran normales, no obstante, las concentraciones de IgM séricas y los títulos de anticuerpos específicos frente a los 14 serotipos de polisacáridos neumocócicos fueron bajos. Los complementos CH50, C3, C4, y la fagocitosis de neutrófilos y la explosión oxidativa fueron normales. La expresión de los receptores toll-like en los monocitos también fue normal. Por lo tanto, hay que descartar que los pacientes adultos que presentan infecciones graves potencialmente mortales y poco comunes no tengan una posible insuficiencia de IgM selectiva primaria.

Palabras clave: Insuficiencia de IgM selectiva. Inmunoinsuficiencia primaria. Infecciones poco comunes. Receptores toll-like. Autoanticuerpos. Aspergilosis. Complemento. Complejos inmunes

Introduction

Selective immunoglobulin (Ig) M deficiency is a rare form of primary immunodeficiency with a reported prevalence of 0.03% to 3% [1]. Selective IgM deficiency can be asymptomatic or present symptomatically with infections caused by encapsulated bacteria and viruses, some

of which can be serious and even life-threatening. These vary from pneumonia to septicemia and meningitis [2-4]. Selective IgM deficiency is a heterogeneous disorder with no known genetic component, and may occur as a primary or a secondary condition. Secondary selective IgM deficiency is often associated with malignant neoplasm or autoimmune diseases [5-8].

Case Description

A 49-year-old previously healthy man with an unremarkable clinical history presented with fevers, chills, malaise, and a generalized petechial rash. He quickly developed septic shock and respiratory failure, and was admitted to the intensive care unit where he was intubated and received multiple vasopressors, aggressive intravenous fluid hydration, and parenteral antibiotics. Multiple blood cultures grew *Streptococcus pneumoniae*. His hospital stay was prolonged and complicated. He suffered acute renal failure requiring dialysis and disseminated intravascular coagulation with deep vein thrombosis of the inferior vena cava and bilateral iliac veins for which an inferior vena cava filter was implanted. He also developed acute infectious purpura fulminans involving over 30% of his body. This progressed from the initial petechial rash to larger confluent ecchymotic areas, some of which became necrotic and gangrenous, and necessitated amputation of his right leg below the knee and multiple excisional débridements and skin grafts. The right leg stump became infected with vancomycin-resistant *Enterococcus faecium* and *Candida*

White blood cell count, $10^3/\mu L$ 15.6 4.0-10.5 Hemoglobin, g/dL 9.3 13.5-16.9 Plateles, $10^3/\mu L$ 96 150-400 Absolute neutrophil count, $10^3/\mu L$ 0.8 0.9-3.3 Absolute neutrophil count, $10^3/\mu L$ 0.5 0.0-0.8 Absolute noncyte count, $10^3/\mu L$ 0.3 0.0-0.54 HIV ELISA Negative Negative Adaptive immunity Lymphocyte subsets, $No./\mu L$ (%) CD3 + CD4 + T cells 1062 (68) 338-1194 (31-61) CD3 + CD4 + T cells 1062 (68) 338-1194 (31-61) CD3 + CD4 + T cells 1062 (68) 0.9-3.7) CD3 + CD4 + T cells 1265 (10) 85-729 (10-38) Ratio of CD4/CD8 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagulutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitigen 213 399 20171-78 728 Mumps antigen 3763 2052-26 495 Candida albicars antigen 4957 132 49-60 917 Tetan		Patient R	esult	Referenc	e Range
Plateles, $10^5\mu$ L 96 150-400 Absolute neutrophil count, $10^5\mu$ L 14 2.0-8.1 Absolute mynhocyte count, $10^5\mu$ L 0.8 0.9-3.3 Absolute monocyte count, $10^5\mu$ L 0.5 0.0-0.8 Absolute monocyte count, $10^5\mu$ L 0.3 0.0-0.54 HIV ELISA Negative Negative Adaptive immunity Lymphocyte subsets, No./µL (%) CD3 + T cells 1265 (81) 619-1847 (62-84) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + T cells 156 (10) 85-729 (10-38) Ratio of CD4/CD8 (6.8) (0.9-3.7) (10.9-3.7) (10.9-3.7) CD3-CD19 + B cells 219 (14) 51-473 (5-26) CD3-CD5 + NK cells 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagulutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21399 2017.17-87 728					
Absolute neutrophil count, $10^3/\mu$ L 14 2.0-8.1 Absolute noncovte count, $10^3/\mu$ L 0.8 0.9-3.3 Absolute onnocyte count, $10^3/\mu$ L 0.8 0.9-3.3 Absolute cosinophil count, $10^3/\mu$ L 0.3 0.0-0.54 HIV ELISA Negative Negative Adaptive immunity Jymphocyte subsets, No./µL (%) 0.5 CD3 + T cells 1265 (81) 619-1847 (62-84) CD3 + CD4 + T cells 1062 (68) 338-1194 (31-61) CD3 + CD4 + T cells 1062 (6.8) (0.9-3.7) (CD3-CD56 + NK cells 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 25 25 25 26 2052-26 495 Candida albicans antigen 3763 2052-26 495 2052-26 495 2052-26 495 20171-78 728 Mumps antigen 3763 2052-26 495 2052-26 495 20171-78 728 Mumps antigen 31244-545-2580 545-2580 56 39 Tuberculin antigen 1324 -545-2580 56 33					
Absolute monocyte count, $10^5/\mu L$ 0.8 0.9-3.3 Absolute monocyte count, $10^5/\mu L$ 0.5 0.0-0.8 Absolute cosinophil count, $10^5/\mu L$ 0.3 0.0-0.54 HIV ELISA Negative Negative Adaptive immunity L_{sprint} Negative CD3 + T cells 1265 (81) 619-1847 (62-84) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + T cells 156 (10) 85-729 (10-38) Ratio of CD4/CD8 (6.8) (0.9-3.7) CD3-CD19 + B cells 219 (14) 51-473 (5-26) CD3-CD56 + NK cells 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 <					
Absolute nonocyte count, $10^3/\mu$ L 0.5 0.0-0.8 Absolute cosinophil count, $10^3/\mu$ L 0.3 0.0-0.54 HIV ELISA Negative Negative Adaptive immunity Lymphocyte subsets, No./ μ L (%) 0.2 CD3 + T cells 1265 (81) 619-1847 (62-84) CD3 + CD4 + T cells 1062 (68) 338-1194 (31-61) CD3 + CD4 + T cells 156 (10) 85-729 (10-38) Ratio of CD4/CD8 (6.8) (0.9-3.7) CD3 - CD5 + NK cells 219 (14) 51-473 (5-26) CD3 - CD5 + NK cells 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 13 129-52 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 104-60 917 74-38 124 Serum immunoglobulins IgA, mg/dL 18 65-263					
Absolute cosinophil count, $10^{5/}\mu L$ 0.3 0.0-0.54 HIV ELISA Negative Negative Adaptive immunity Lymphocyte subsets, No./ μL (%) CD3 + T cells 1265 (81) 619-1847 (62-84) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + CD8 + T cells 156 (10) 85-729 (10-38) Ratio of CD4/CD8 (6.8) (0.9-3.7) (CD3-CD19 + B cells 219 (14) 51-473 (5-26) CD3-CD56 + NK cells 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 733 249-60 917 Tetanus toxoid 907 6092-94 539 7 13 249-545-2580 Serum immunoglobulins IgA, mg/dL 18 65-263 194, mg/dL					
HIV ELISA Negative Negative Adaptive immunity Lymphocyte subsets, No./µL (%) 1265 (81) 619-1847 (62-84) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + T cells 156 (10) 85-729 (10-38) Ratio of CD4/CD8 (6.8) (0.9-3.7) (5-26) CD3-CD56 + NK cells 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokewed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins [gd, mg/dL 124 10-150 [gd, mg/dL 124 10-150 IgG, mg/dL 124 <td< td=""><td></td><td></td><td></td><td colspan="2"></td></td<>					
Adaptive immunity Lymphocyte subsets, $No./\mu L$ (%) CD3 + T cells 1265 (81) 619-1847 (62-84) CD3 + T cells 1062 (68) 338-1194 (31-61) CD3 + CD4 + T cells 156 (10) 85-729 (10-38) Ratio of CD4/CD8 (6.8) (0.9-3.7) (D3-3CD19 + B cells 219 (14) 51-473 (5-26) CD3 - CD5 + NK cells 219 (14) 51-473 (5-26) (C3-20) In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 (Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins Ige, mg/dL 124 10-150 196 IgG, mg/dL 124 10 1323-548 2463 27-134 IgeG,					
$Lymphocyte subsets, No./\muL (%) \\ CD3 + T cells 1265 (81) 619-1847 (62-84) \\ CD3 + T cells 1062 (68) 338-1194 (31-61) \\ CD3 + CD4 + T cells 156 (10) 85-729 (10-38) \\ (0.9-3.7) \\ CD3 - CD4 - T cells 219 (14) 51-473 (5-26) \\ CD3 - CD56 + NK cells 62 (4) 12-349 (1-17) \\ \hline N vitro lymphocyte proliferative response (cpm) \\ Phytohemagglutinin 198 871 114 881-289 206 \\ Concanavalin 174 387 131 199-252 925 \\ Pokeweed mitogen 21 399 20 171-78 728 \\ Mumps antigen 3763 2052-26 495 \\ Candida albicans antigen 4957 13 249-60 917 \\ Tetanus toxoid 907 6092-94 539 \\ Tuberculin antigen 1324 -545-2580 \\ \hline Serum immunoglobulins \\ IgM, mg/dL 18 65-263 \\ IgA, mg/dL 1060 664-378 \\ IgE, mg/dL 1060 664-378 \\ IgE, mg/dL 219 123-548 \\ IgG, mg/dL 28 27-134 \\ IgG4, mg/dL 9 8-88 \\ \hline Autoantibodies \\ Anti-dsDNA antibody Negative Ne$	HIVELISA	negauve		Inegative	
$\begin{array}{c} CD3 + T \ cells \\ CD3 + CD4 + T \ cells \\ CD3 + CD4 + T \ cells \\ CD3 + CD4 + T \ cells \\ CD3 + CD8 + T \ cells \\ CD3 + CP8 + T \ cells \\ CD3 +$					
$\begin{array}{c} \text{CD3} + \text{CD4} + \text{T cells} & 1062 & (68) & 338-1194 & (31-61) \\ \text{CD3} + \text{CD8} + \text{T cells} & 156 & (10) & 85-729 & (10-38) \\ \text{Ratio of CD4/CD8} & (6.8) & (0.9-3.7) \\ \text{CD3-CD19} + \text{B cells} & 219 & (14) & 51-473 & (5-26) \\ \text{CD3-CD56} + \text{NK cells} & 62 & (4) & 12-349 & (1-17) \\ \hline \textit{In vitro lymphocyte proliferative response (cpm)} \\ \text{Phytohemagglutinin} & 198 871 & 114 881-289 206 \\ \text{Concanavalin} & 174 387 & 131 199-252 925 \\ \text{Pokeweed mitogen} & 21 399 & 20 171-78 728 \\ \text{Mumps antigen} & 3763 & 2052-26 495 \\ \hline \textit{Candida albicans antigen} & 4957 & 13 249-60 917 \\ \text{Tetanus toxoid} & 907 & 6092-94 539 \\ \text{Tuberculin antigen} & 1324 & -545-2580 \\ \hline \hline \textit{Serum immunoglobulins} \\ \text{IgM, mg/dL} & 18 & 65-263 \\ \text{IgA, mg/dL} & 1066 & 694-1618 \\ \text{IgG1, mg/dL} & 1066 & 694-1618 \\ \text{IgG1, mg/dL} & 219 & 123-548 \\ \text{IgG3, mg/dL} & 219 & 123-548 \\ \text{IgG3, mg/dL} & 219 & 123-548 \\ \text{IgG4, mg/dL} & 9 & 8-88 \\ \hline \hline \textit{Autoantibodies} \\ \hline \textit{Anti-dsDNA antibody} & \text{Negative} & \text{Negative} \\ \hline \textit{Anti-Smith} & \text{Negative} & \text{Negative} \\ \textit{Anti-Smith} & \text{Negative} & \text{Negative} \\ \textit{Lymphocyte apoptosis (% cells)^{\mu}} \\ \hline \end{array}$					
$\begin{array}{cccccccccccccccccccccccccccccccccccc$					
Ratio of CD4/CD8 (6.8) (0.9-3.7) CD3-CD19 + B cells 219 (14) $51-473$ (5-26) CD3-CD56 + NK cells 62 (4) 12-349 (1-17) <i>In vitro lymphocyte proliferative response (cpm)</i> Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 <i>Candida albicans</i> antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins Ige, mg/dL 18 IgG, mg/dL 180 68-378 IgG, mg/dL 28 27-134 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies Negative Negative ANA 1:160 (Nucleolar) <1:40					
$\begin{array}{c} \text{CD3-CD19 + B cells} & 219 & (14) & 51-473 & (5-26) \\ \text{CD3-CD56 + NK cells} & 62 & (4) & 12-349 & (1-17) \\ \hline \textit{In vitro lymphocyte proliferative response (cpm)} \\ \text{Phytohemagglutinin} & 198 871 & 114 881-289 206 \\ \text{Concanavalin} & 174 387 & 131 199-252 925 \\ \text{Pokeweed mitogen} & 21 399 & 20 171-78 728 \\ \text{Mumps antigen} & 3763 & 2052-26 495 \\ \textit{Candida albicans antigen} & 4957 & 13 249-60 917 \\ \text{Tetanus toxoid} & 907 & 6092-94 539 \\ \text{Tuberculin antigen} & 1324 & -545-2580 \\ \hline \textit{Serum immunoglobulins} \\ \text{IgA, mg/dL} & 18 & 65-263 \\ \text{IgA, mg/dL} & 124 & 10-150 \\ \text{IgG, mg/dL} & 10660 & 694-1618 \\ \text{IgG1, mg/dL} & 219 & 123-548 \\ \text{IgG3, mg/dL} & 219 & 123-548 \\ \text{IgG3, mg/dL} & 28 & 27-134 \\ \text{IgG4, mg/dL} & 9 & 8-88 \\ \hline \textit{Autoantibodies} \\ \text{ANA} & 1:160 (Nucleolar) & <1:40 \\ \text{Anti-dsDNA antibody} & \text{Negative} & \text{Negative} \\ \text{RF} & \text{Negative} & \text{Negative} \\ \text{Negative} \\ \text{Negative} & \text{Negative} \\ \text{Negative} \\ \text{Negative} \\ \text{Negative} \\ \text{Negative} \\ \text{Negative} \\ Negat$		156		85-729	
CD3-CD56 + NK cells 62 (4) 12-349 (1-17) In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins IgM, mg/dL 18 65-263 IgA, mg/dL 180 68-378 IgG1, mg/dL 1060 694-1618 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies Negative Negative ANA 11:160 (Nucleolar) <1:40					
In vitro lymphocyte proliferative response (cpm) Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins 1 18 65-263 IgA, mg/dL 18 65-263 1 IgA, mg/dL 124 10-150 1060 694-1618 IgG1, mg/dL 1060 694-1618 1 126-239-1083 IgG2, mg/dL 219 123-548 1 123-548 IgG3, mg/dL 28 27-134 1 126-488 Autoantibodies - - - - ANA 1:160 (Nucleolar) <1:40					
Phytohemagglutinin 198 871 114 881-289 206 Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins 124 10-150 IgG, mg/dL 18 65-263 IgG, mg/dL 124 10-150 IgG, mg/dL 124 10-150 IgG, mg/dL 219 123-548 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies ANA 1:160 (Nucleolar) ANA 1:160 (Nucleolar) <1:40	CD3-CD56 + NK cells	62	(4)	12-349	(1-17)
Concanavalin 174 387 131 199-252 925 Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins IgA, mg/dL 18 65-263 IgA, mg/dL 180 68-378 IgE, mg/dL 104 10060 694-1618 IgG1, mg/dL 219 123-548 123-548 IgG2, mg/dL 21 28 27-134 IgG4, mg/dL 9 8-88 9 Autoantibodies ANA 1:160 (Nucleolar) <1:40	In vitro lymphocyte proliferative response (cpm)				
Pokeweed mitogen 21 399 20 171-78 728 Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins 18 65-263 IgA, mg/dL 18 68-378 IgE, mg/dL 124 10-150 IgG, mg/dL 1060 694-1618 IgG1, mg/dL 28 27-134 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies Negative Negative ANA 1:160 (Nucleolar) <1:40					
Mumps antigen 3763 2052-26 495 Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 6092-94 539 Tuberculin antigen 1324 -545-2580 Serum immunoglobulins IgM, mg/dL 18 65-263 IgA, mg/dL 180 68-378 IgE, mg/dL 124 10-150 IgG, mg/dL 1060 694-1618 IgG1, mg/dL 219 123-548 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies ANA 1:160 (Nucleolar) <1:40					
Candida albicans antigen 4957 13 249-60 917 Tetanus toxoid 907 $6092-94 539$ Tuberculin antigen 1324 $-545-2580$ Serum immunoglobulins IgM, mg/dL 18 $65-263$ IgA, mg/dL 180 $68-378$ IgE, mg/dL 124 10-150 IgG, mg/dL 1060 $694-1618$ IgG1, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies 496 8-88 Autoantibody Negative Negative ANA 1:160 (Nucleolar) <1:40					
Tetanus toxoid 907 $6092-94539$ Tuberculin antigen 1324 $-545-2580$ Serum immunoglobulins IgM, mg/dL 18 $65-263$ IgA, mg/dL 180 $68-378$ IgE, mg/dL 1060 IgG, mg/dL 1060 $694-1618$ IgG1, mg/dL 239-1083 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 8 9 8-88 Autoantibodies					
Tuberculin antigen 1324 -545-2580 Serum immunoglobulins IgM, mg/dL 18 65-263 IgA, mg/dL 180 68-378 IgE, mg/dL 124 10-150 IgG, mg/dL 1060 694-1618 IgG1, mg/dL 660 239-1083 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies 1:160 (Nucleolar) <1:40					
Serum immunoglobulins 18 65-263 IgA, mg/dL 180 68-378 IgE, mg/dL 124 10-150 IgG, mg/dL 1060 694-1618 IgG1, mg/dL 660 239-1083 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies 1:160 (Nucleolar) <1:40					
IgM, mg/dL 18 65-263 IgA, mg/dL 180 68-378 IgE, mg/dL 124 10-150 IgG, mg/dL 1060 694-1618 IgG1, mg/dL 660 239-1083 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies 7 1:160 (Nucleolar) <1:40	Tuberculin antigen	1324		-545-2	2580
IgA, mg/dL 180 $68-378$ IgE, mg/dL 124 $10-150$ IgG, mg/dL 1060 694.1618 IgG1, mg/dL 660 $239-1083$ IgG2, mg/dL 219 $123-548$ IgG3, mg/dL 28 $27-134$ IgG4, mg/dL 9 $8-88$ Autoantibodies ANA 1:160 (Nucleolar) $<1:40$ Anti-dsDNA antibody Negative Negative ANCA Negative Negative RF Negative Negative Anti-Smith Negative Negative Anti-RNP Negative Negative Lymphocyte apoptosis (% cells) ^a					
IgE, mg/dL 124 10-150 IgG, mg/dL 1060 694-1618 IgG1, mg/dL 660 239-1083 IgG2, mg/dL 219 123-548 IgG3, mg/dL 28 27-134 IgG4, mg/dL 9 8-88 Autoantibodies ANA 1:160 (Nucleolar) <1:40					
IgG, mg/dL1060 $694-1618$ IgG1, mg/dL 660 $239-1083$ IgG2, mg/dL 219 $123-548$ IgG3, mg/dL 28 $27-134$ IgG4, mg/dL 9 $8-88$ AutoantibodiesANA $1:160$ (Nucleolar) $<1:40$ Anti-dsDNA antibodyNegativeNegativeANCANegativeNegativeRFNegativeNegativeAnti-SmithNegativeNegativeAnti-RNPNegativeNegativeLymphocyte apoptosis (% cells) ^a 4000					
IgG1, mg/dL 660 $239-1083$ IgG2, mg/dL 219 $123-548$ IgG3, mg/dL 28 $27-134$ IgG4, mg/dL 9 $8-88$ AutoantibodiesANA $1:160$ (Nucleolar) $<1:40$ Anti-dsDNA antibodyNegativeNegativeANCANegativeNegativeRFNegativeNegativeAnti-SmithNegativeNegativeAnti-RNPNegativeNegativeLymphocyte apoptosis (% cells) ^a 460					
IgG2, mg/dL219123-548IgG3, mg/dL2827-134IgG4, mg/dL98-88AutoantibodiesANA1:160 (Nucleolar)Anti-dsDNA antibodyNegativeANCANegativeRFNegativeAnti-SmithNegativeAnti-RNPNegativeLymphocyte apoptosis (% cells) ^a					
IgG3, mg/dL2827-134IgG4, mg/dL98-88AutoantibodiesANA1:160 (Nucleolar)Anti-dsDNA antibodyNegativeANCANegativeANCANegativeRFNegativeAnti-SmithNegativeAnti-RNPNegativeLymphocyte apoptosis (% cells) ^a					
IgG4, mg/dL98-88AutoantibodiesANA1:160 (Nucleolar)<1:40					
Autoantibodies ANA 1:160 (Nucleolar) <1:40					
ANA1:160 (Nucleolar)<1:40Anti-dsDNA antibodyNegativeNegativeANCANegativeNegativeRFNegativeNegativeAnti-SmithNegativeNegativeAnti-RNPNegativeNegativeLymphocyte apoptosis (% cells) ^a	IgG4, mg/dL	9		8-8	8
Anti-dsDNA antibodyNegativeNegativeANCANegativeNegativeRFNegativeNegativeAnti-SmithNegativeNegativeAnti-RNPNegativeNegativeLymphocyte apoptosis (% cells) ^a	Autoantibodies				
ANCA Negative Negative RF Negative Negative Anti-Smith Negative Negative Anti-RNP Negative Negative					
RF Negative Negative Anti-Smith Negative Negative Anti-RNP Negative Negative	Anti-dsDNA antibody				
Anti-Smith Anti-RNP Negative Negative Negative Negative Lymphocyte apoptosis (% cells) ^a					
Anti-RNP Negative Negative Lymphocyte apoptosis (% cells) ^a Image: Comparison of the second	RF				
Lymphocyte apoptosis (% cells) ^a	Anti-Smith	Negativ	ve		
	Anti-RNP	Negativ	ve	Nega	tive
	<i>Lymphocyte apoptosis (% cells)</i> ^a				
		18%		10%-2	20%

Abbreviations:ANA, antinuclear antibodies; ANCA, antineutrophil cytoplasmic antibodies; cpm, counts per minute; dsDNA, double-stranded DNA; ELISA, enzyme-linked immunosorbent assay; HIV, human immunodeficiency virus; lg, immunoglobulin; RF, rheumatoid factor; RNP, ribonucleoprotein antibodies

^a Apoptosis was measured by Annexin V binding

	Patient Result		Reference Range	
Adaptive immunity (continued)				
Specific antibodies				
Tetanus toxoid antibody (IgG), IU/mL	0.22		>1	
Pneumococcal antibody (IgG µg/mL)	Pre ^a	Post ^b		
Serotype 1	0.34	0.65		
Serotype 3	0.11	0.12		
Serotype 4	0.19	0.18		
Serotype 5	0.59	0.98		
Serotype 6B	0.56	0.46		
Serotype 7F	0.21	0.60		
Serotype 8	0.12	0.38		
Serotype 9N	0.13	0.40		
9Serotyope V	0.13	0.32		
Serotype 12F	0.49	0.50		
14, µg/mL	0.66	1.00		
18C, µg/mL	0.23	0.68		
19F, µg/mL	0.45	0.92		
23F, µg/mL	0.07	0.42		
Innate immunity				
C3, mg/dL	128		88-201	
C4, mg/dL	32		16-47	
CH50 U/mL	340		101-300	
Raji immune complex µgE/mL	9		0-25	
C1q binding assay µgE/mL	2.4 >20		0-3.9 0-0.7	
C reactive protein, mg/dL Reactive oxygen species (index)	2.2		2-18	
Phagocytosis assay (% phagocytosis)	38.4		25-45	
TLR2 + CD14 + monocytes	74.4		19-76	

Table. Immunologic Studies (continued)

Abbreviations: Ig, immunoglobulin; TLR2, toll-like receptor 2

^a Pre = IgG > 1 μ g/mL

^b Post = pre x4

albicans, and an additional course of antibiotics was necessary. Five weeks after presentation, necrotic tissue was observed in his nostrils. He was diagnosed with acute invasive fungal sinusitis following nasal biopsy and right maxillary aspirate cultures, which grew *Aspergillus funigatus*: subsequent débridement was performed. Physical examination revealed post-surgical changes in his nose and the amputation stump. Skin examination revealed multiple erythematous and necrotic rashes on his arms and legs.

Lymphocyte subsets and toll-like receptor (TLR) expression on monocytes were determined by multicolor flow cytometry using direct fluorochrome-conjugated antibodies against CD3, CD4, CD8, CD19, CD16/CD56, CD14/TLR2, and isotype controls. Lymphocyte proliferation was measured by culturing mononuclear cells (2×10^5 /well) in triplicate in round-bottom tissue culture plates at 37°C in the presence or absence of optimal concentrations of mitogens and antigens and by incorporation of 3H-thymidine. Data were expressed as net counts per minute.

Phagocytosis of neutrophils was performed on 100 μ L of whole blood, which was incubated with phycoerythrinconjugated yeast or unlabeled yeast for 15 minutes at room temperature. Samples were incubated with 2 mL of fluorescenceactivated cell sorting (FACS) lysing solution for 15 minutes and washed with 2 mL of phosphate-buffered saline. Flow cytometry analysis was performed with a FACSCalibur flow cytometer (Becton Dickinson, San Jose, California, USA). Reactive oxygen species (ROS) as a measure of oxidative burst was generated on 100 μ L of whole blood, which was incubated at 37°C with oxidation-dependent fluorescence dihydrorhodamine 123 (2.5 μ g/mL) for 15 minutes, then stimulated with phorbol-12-myristate-13 acetate (2.5 μ g/mL) for an additional 15 minutes. The samples were acquired on the FACSCalibur. All flow cytometry data were analyzed using Simulset (Becton Dickinson, San Jose, California, USA) software.

Comprehensive immunologic evaluation is presented in the Table. The white blood cell count was high with an elevated neutrophil percentage. T cells, T cell subsets, B cells, and natural killer cells were within normal limits. Lymphocyte transformation of phytohemagglutinin, concanavalin A, pokeweed mitogen, mumps antigen, and purified protein derivative antigen was normal. However, low lymphocyte transformation to *C albicans* and tetanus toxoid was observed, suggesting a T cell functional defect. The patient had normal quantitative serum IgG and IgG subclasses, IgA, and IgE. However, serum IgM levels were low at 18 mg/dL and the patient failed to make an antibody

response to all 14 pneumococcal polysaccharide serotypes (even in the presence of *S pneumoniae* sepsis) and to tetanus toxoid. Furthermore, the patient made no antibodies following Pneumovax vaccination. Repeated determinations showed low levels of IgM, and the patient remained IgM-deficient even after 1 year from the initial diagnosis. Neutrophil phagocytic capacity and generation of ROS were normal. TLR2 on monocytes was normal. C-reactive protein (CRP) was markedly elevated. Circulating immune complexes were negative. The antinuclear antibody test result was positive, although the results of testing for anti-dsDNA antibody, antineutrophil cytoplasmic antibody, rheumatoid factor, anti-Smith antibody, and antiribonucleoprotein antibody were negative.

Discussion

Primary selective IgM deficiency is a rare disorder in children and has no known genetic component. However, familial cases [4] and autosomal dominant inheritance [9] have been suggested. Primary selective IgM deficiency in children may present with severe life-threatening infections, whereas in adults it is usually associated with autoimmune diseases and malignant neoplasm [5-8]. A few cases of possible primary selective IgM deficiency in adults with no evidence of autoimmunity or malignant neoplasm have been reported [9,10]. These patients usually present with mild infections. Our patient, who was previously healthy with no evidence of autoimmunity or neoplasm, represents a possible case of primary selective IgM deficiency in adults with severe life-threatening and unusual infections. Numbers of B cells, especially with surface IgM, are generally normal, high, or decreased [5,9,10]. Our patient also had normal proportions of CD19+ B cells. Although levels of IgG and IgG subclasses were normal, our patient had decreased specific IgG antibody response to tetanus toxoid and to all 14 serotypes of pneumococcal polysaccharides. Guill et al [11] also reported decreased specific antibody response to tetanus toxoid and pneumococcal polysaccharide, and decreased IgM response to immunization with ϕ X174 in children with selective IgM deficiency. Proportions and numbers of CD4+ T cells and CD8+T cells and CD4+/CD8+T cell ratios have been reported to be normal, low, or high [5,9]. In our patient, the CD4+/CD8+ T cell ratio was abnormally high.

The pathogenesis of primary selective IgM deficiency is unknown. A number of defects have been reported, including intrinsic B cell defect in plasma cell differentiation [6], increased T cell suppressor activity, which may be specific to IgM isotype [5,7,9] or isotype-nonspecific [9], and decreased helper T cell activity [10]. This suggests that selective IgM deficiency is a heterogeneous disorder, which requires further studies to elucidate the predominant mechanisms involved in its pathogenesis.

Our patient is unique in that he had severe infections with 2 organisms, *S pneumoniae* and *A fumigatus*, both containing polysaccharide antigens. *S pneumoniae* is an important pathogen in humans, and both adaptive and innate immune mechanisms provide protection from infection. Although protective anticapsular antibodies can be produced following immunization, the innate immune responses are clearly important in controlling infections in the nonimmune host. Our patient failed to produce protective anticapsular antibodies against any of the 14 serotypes tested. Several components of the innate immune system that play a role in defense against *S pneumoniae* include CRP and signaling via TLRs. CRP provides protection against *S pneumoniae* in both a complement-dependent and complement-independent manner. Our patient had increased CRP and complement levels were normal. Therefore, it appears unlikely that a defect in a CRP-dependent mechanism was responsible for *S pneumoniae* sepsis. TLRs are important in initiating innate responses to a wide variety of pathogens [12]: TLR2 recognizes lipoteichoic acid, lipoarabinomannan, and lipopeptides, whereas TLR4 recognizes lipopolysaccharide. In our patient, TLR2 expression on CD14+ monocytes was normal.

The first line of defense against *A fumigatus* is provided by innate immunity, predominantly by macrophages via secretion of tumor necrosis factor- α (TNF- α). Various mechanisms of TNF– α -mediated resistance against *A fumigatus* include upregulation of phagocytosis and augmentation of antibodydependent cytotoxicity and of oxidative burst to kill A fumigatus [13,14]. Our patient had normal polymorphonuclear cell phagocytosis. Although a role for specific antibodies in defense against *A fumigatus* has not been described, it is possible that specific antibody responses against the polysaccharide antigen galactomannan may play a role in late defense response, and, therefore, a polysaccharide antibody defect in our patient may be responsible for the development of invasive aspergillosis.

The take-home messages from this patient are as follows: (a) adult patients with serious *S pneumoniae* infection should be investigated for possible selective IgM deficiency; (b) both children and adults with possible primary selective IgM deficiency are susceptible to life-threatening and often unusual infections; and (c) taking into consideration the history of serious and unusual infection in this patient with selective IgM deficiency and various host defense mechanisms against these organisms, patients with selective IgM deficiency should also be investigated for innate immune response.

References

- Cassidy JT, Nordby GL. Human serum immunoglobulin concentrations: prevalence of immunoglobulin deficiencies. J. Allergy Clin Immunol. 1975;55:35-48.
- Faulk WP, Kiyasu WS, Cooper MD, Fudenberg HH. Deficiency of IgM. Pediatrics. 1971;47:399-404.
- Hobbs JR, Milner RDG, Watt PJ. Gamma-M deficiency predisposing to meningococcal septicaemia. BMJ. 1967;4:583-6.
- Ross IN. Thompson RA. Severe selective IgM deficiency. J Clin Pathol. 1976;29:773-7.
- Inoue T, Okumura Y, Shirahama M, Ishibashi H, Kashiwagi S, Okubo H. Selective partial IgM deficiency: Functional assessment of T and B lymphocytes in vitro. J Clin Immunol. 1986;6:130-5.
- Goldstein MF, Goldstein AL, Dunsky EH, Dvorin DJ, Belecanech GA, Shamir K. Selective IgM deficiency: retrospective analysis of 36 adult patients with review of the literature. Ann Allergy Asthma Immunol. 2006;97:717-30.
- Takeuchi T, Nakagawa T, Maeda Y, Hirano S, Sasaki-Hayashi M, Makino S, Shimizu A. Functional defect of B lymphocytes in a

patient with selective IgM deficiency associated with systemic lupus erythematosus. Autoimmunity. 2001;34:115-22.

- 8. Vogelzang NJ, Corwin H, Finlay JL, Pellettiere EV 2nd, Luskin AT, Di Camelli RF, Hong R. Clear cell sarcoma and selective IgM deficiency: a case report. Cancer. 1982;49:234-8.
- 9. Ohno T, Inaba M, Kuribayashi K. Selective IgM deficiency in adults: phenotypically and functionally altered profiles of peripheral blood lymphocytes. Clin Exp Immunol. 1987;68:630-7.
- De la Concha EG, Garcia-Rodriguez MC, Zabay JM. Functional assessment of T and B lymphocytes in patients with selective IgM deficiency. Clin Exp Immunol. 1982;49:670-6.
- Guill MF, Brown DA, Ochs HD, Pyun K, Moffitt J. IgM deficiency: clinical spectrum and immunological assessment. Ann Allergy. 1989;62:547-52.
- 12. Akira S and Takeda K. Toll-like receptor signaling. Nature Rev Immunol. 2004;4:499-511.
- 13. Roilides E, Dimitriadou-Georgiadou A, Sein T, Kadilt-soglou I, Walsh TJ. Tumor necrosis factor alpha enhances antifungal activities

of polymorphonuclear and mononuclear phagocytes against Aspergillus fumigatus. Infect Immun. 1998;66:5999-6003.

 Ferrante A, Nandoskar M, Walz A, Goh DHB, Kowanko IC. Effects of tumor necrosis factor alpha and interleukin-1 alpha and beta on human neutrophil migration, respiratory burst and degranulation. Int Arch Appl Immunol. 1988;86:82-91.

Manuscript received July 25, 2007; accepted for publication October 10, 2007.

Sudhir Gupta, MD, PhD

Medical Sciences I, C240 University of California, Irvine Irvine, CA 92697, United States Phone: (949) 824-5818 Fax: (949) 824-4362 E-mail: sgupta@uci.edu